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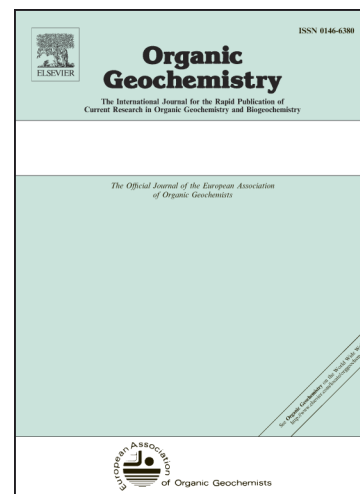
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ABSTRACT

Long-chain alkenones (LCAs) have been used for decades to reconstruct quantitative sea-surface temperature records, but they also have a great potential for paleotemperature reconstructions in lacustrine settings. Here, we investigated how the presence and abundance of LCAs in surface sediments from 106 lakes varied with environmental conditions in lakes of the Northern Great Plains (Canadian Prairies) in southern Saskatchewan, Canada. Consistent with preliminary research, we found LCAs in 55% of surveyed lakes, with mean concentrations of 143 $\mu\text{g/g}$ dry sediment, but very high concentrations (up to 2.3 mg/g dry sediment) in 7% of lakes. Statistical analyses indicate that salinity and stratification play key roles in determining LCA presence and abundance supporting previous research in Spain and the northern Great Plains, USA. Overall, the alkenone unsaturation index (U_{37}^K) was not correlated significantly with average summer water temperature, probably because the timing of maximum LCA production occurs during the spring season. We conclude that improved seasonal sampling is required within the study lakes to better identify the timing and habitat of haptophyte production, and allow development of environmental temperature reconstruction tools.

Keywords: alkenones, lakes, haptophyte, salinity, stratification, U_{37}^K .

1. Introduction

Long-chain alkenones (LCAs) are C_{35} – C_{42} di-, tri- and tetra-unsaturated methyl and ethyl ketones whose biosynthesis in modern oceans is restricted to a few unicellular haptophyte algae of the order Isochrysidales, including the calcifying species (coccolithophores) *Emiliania huxleyi* and *Gephyrocapsa oceanica* (Volkman et al., 1980, 1995; Marlowe et al., 1984), and the coastal non-calcifying species *Ruttenella lamellosa* (formerly *Chrysotila lamellosa*) and *Isochrysis galbana* (Marlowe et al., 1984, 1990). The degree of unsaturation of C_{37} alkenones has been shown to vary as a function of the environmental temperature experienced during haptophyte growth (Brassell et al., 1986; Prahl and Wakeham, 1987), and the so-called alkenone unsaturation indices (U_{37}^K , $U_{37}^{K'}$) have been developed in both laboratory cultures and marine surface sediments as proxies for sea surface temperature (Prahl et al., 1988; Müller et al., 1998; Conte et al.,

2006). During the last three decades, the $U_{37}^{K'}$ proxy has been successfully applied to reconstruct past marine temperature records at various sites and timescales (e.g., Rosell-Melé et al., 1995; Müller et al., 1998; Pahnke and Sachs, 2006).

More recently, LCAs have been reported worldwide in continental lakes (e.g., Cranwell, 1985; Zink et al., 2001; Chu et al., 2005; Longo et al., 2016). Unlike the open oceans, where the LCA production is believed to be limited to *E. huxleyi* and *G. oceanica*, genomic analyses from lacustrine sediments and particulate organic matter in lake water have shown the existence of multiple LCA-producing haptophyte species within and among the lakes (e.g., Coolen et al., 2004; D'Andrea et al., 2006; Theroux et al., 2010; Toney et al., 2012). Theroux et al. (2010) identified three distinct phylogenetic groups within the order Isochrysidales. Group I (aka “Greenland phylotype”: D'Andrea et al., 2006) seems to include haptophyte species restricted to freshwater and oligohaline lakes, whereas Group II includes *R. lamellosa* and *I. galbana*, as well as related species found in brackish waters and saline lakes. Group III includes the marine producers *E. huxleyi* and *G. oceanica*. In principle, the presence of multiple species complicates development of biogeochemical metrics of past lake temperatures as the relationship between temperature and the alkenone unsaturation index could differ between haptophyte species (Theroux et al., 2010; Toney et al., 2010; Randlett et al., 2014).

Moreover, the environmental parameters controlling alkenone distribution in inland waters remain unclear as LCAs have been found in freshwater (e.g., Zink et al., 2001; D'Andrea and Huang, 2005; Longo et al., 2013, 2016; D'Andrea et al., 2016) and saline lakes (e.g., Pearson et al., 2008; Liu et al., 2008, 2011; Toney et al., 2010; Zhao et al., 2014; Song et al., 2016), with ionic composition varying from sulfate-rich (e.g., Pearson et al., 2008; Toney et al., 2010) to carbonate-rich solutions (e.g., Chu et al., 2005; D'Andrea and Huang, 2005). Salinity has been suggested as an important factor on the occurrence and concentration of LCAs (D'Andrea and Huang, 2005; Lopez et al., 2005; Pearson et al., 2008; Toney et al., 2010; Liu et al., 2011; Chivall et al., 2014), with thresholds of salinity (< 3 g/L and > 100 g/L) considered unsuitable for LCA producers to prosper (Liu et al., 2011; Song et al., 2016). In addition, seasonality of environmental variables (e.g., temperature, salinity, nutrients), especially in high latitude regions where lakes are ice covered during winter, may have a strong effect on phytoplankton succession (e.g., Lampert and Sommer,

1997; Wetzel, 2001; Korkonen et al., 2017). In fact, lacustrine haptophytes experience sudden increase in abundance (bloom events), likely linked to seasonal changes in nutrient loading and irradiance levels (Toney et al., 2010; D'Andrea et al., 2011; Huguet et al., 2011; Theroux, 2012).

Despite these potential complications, LCAs have been demonstrated to vary with lake temperature in some sites, and a number of calibrations of the U_{37}^K , $U_{37}^{K'}$, and $U_{37}^{K''}$ indices have been established in cultures (e.g., Sun et al., 2007; Toney et al., 2012; Theroux et al., 2013; Zheng et al., 2016) and in situ in freshwater (e.g., Zink et al., 2001; D'Andrea et al., 2011, 2016; Theroux et al., 2013; Longo et al., 2016) and saline lakes (e.g., Chu et al., 2005; Toney et al., 2010; Zheng et al., 2016). Together, these findings suggest that quantitative reconstruction of environmental temperature is possible.

The development of LCAs as metrics for past lacustrine temperatures may be important for the Great Plains of North America. This region is sensitive to temperature and moisture changes and often experiences prolonged periods of drought and floods that have major impacts on agriculture production and ecosystem function. Although lacustrine proxies for past droughts and pluvial intervals have been developed (e.g., diatom- and ostracod-inferred salinity; e.g., Cywinska and Delorme, 1995; Michels et al., 2007), there are few unambiguous proxies of past temperature in inland waters before the instrumental period. Quantitative paleotemperature reconstructions are thus crucial to help decipher the climate mechanisms responsible for drought occurrences. Studies from the northern Great Plains, USA (Theroux et al., 2010; Toney et al., 2010, 2012), suggest that LCA and haptophyte distributions may be suitable for quantitative temperature reconstruction, particularly at well-studied sites. Toney et al. (2011) reported the presence of LCAs in 13 lakes in the Canadian Prairies, southern Saskatchewan, where the development of an alkenone paleothermometer is particularly promising, because there appears to be only one haptophyte species present based on the LCA distribution. However, the low sample size ($n = 19$) was insufficient to conclusively determine which environmental parameters control the presence of LCA-producing haptophytes, or to derive a temperature calibration based on lacustrine surface sediments.

Here, we assess LCA presence and distributions in surface sediments collected from a broader survey of 87 additional lakes in the Canadian Prairies. Water chemistry data measured at the time of sampling for each lake allows us to better statistically constrain the environmental parameters favouring LCA presence and

concentration in these 106 lakes. We also investigated the relationship between LCA and environmental temperature to further develop potential applications of an alkenone unsaturation index, such as U_{37}^K .

2. Methods

2.1. Sites and sampling

Between July 29th and August 30th 2013, 106 lakes from the Canadian Prairies (southern Saskatchewan, Canada) (Fig. 1) were sampled during a field campaign for studies on modern drought risks and ecosystem sustainability. These lakes occur in the sub-humid (north east) to the semi-arid (south-west) areas of the Canadian Prairies, a region where evaporation exceeds precipitation by 20–60 cm/y and lakes are sustained by the influx of snowmelt and groundwater (Pham et al., 2008, 2009). Climate data for the study region, obtained from the WorldClim database, indicate mean (\pm SD) air temperatures and precipitation of; 2.5 ± 1.0 °C and 27.9 ± 2.5 mm in spring, 16.9 ± 0.7 °C and 56.9 ± 6.0 mm in summer, 3.6 ± 0.8 °C and 23.6 ± 4.1 mm in autumn and -14.2 ± 2.0 °C and 16.9 ± 1.9 mm in winter. Survey lakes span a wide spatial extent of 4° latitudinal gradient (Fig. 1) and a large range of salinity from fresh (0.1 g/L) to hypersaline (102 g/L), with diverse ionic composition.

In situ lake parameters, including salinity, conductivity, pH, depth and temperature, were measured at 0.5 m depth intervals (lakes < 10 m depth) or 1 m depth intervals (lakes > 10 m depth) for the entire water column using a YSI Pro Plus meter (YSI Inc., Yellow Springs, Ohio, USA) following Pham et al. (2008). Major ions including sodium (Na^+), chloride (Cl^-), calcium (Ca^{2+}), magnesium (Mg^{2+}), sulfate (SO_4^{2-}), bicarbonate (HCO_3^-), and carbonate (CO_3^{2-}) were determined for each lake and analysed at the University of Alberta Water Chemistry Laboratory using standard procedures (Stainton et al., 1977). Water chemistry, especially surface salinity, of saline lakes may undergo significant changes over the seasonal cycles, and our in situ, one-time measurements of salinity may not capture the seasonal variation of ionic content in these lakes (Pham et al., 2009). However, while extremely dry years lead to spatially coherent increases in lake salinity from spring to later summer in the Canadian prairie lakes (Pham et al., 2009), there is little evidence

of progressive increases in salinity in years with elevated precipitation despite a net precipitation deficit (Pham et al. 2009), such as those which preceded our 2013 survey. These patterns are described in detail as part of the Energy-mass flux framework, outlined in Leavitt et al. (2009). Thus, it is likely that salinity does not significantly change from spring to summer in most of our survey lakes.

Surface sediments were collected from each lake using an Ekman grab sampler. Subsamples of sediments (top cm of the collected sediments) were freeze-dried at 0.01 Pa for 36 h before use for analyses of organic constituents.

2.2. LCA analyses

Freeze-dried sediments were homogenized and extracted with dichloromethane (DCM):methanol (MeOH) (9:1, v:v) using a Dionex model ASE350 accelerated solvent extractor. Following evaporation of the solvent, the total lipid extracts were separated into neutral and acid fractions by elution through a LC-NH₂ SPE column using DCM:isopropyl alcohol (1:1, v:v) followed by ether with 4% acetic acid (v:v) as eluents, respectively. The neutral fractions were further separated into four fractions of increasing polarity by chromatography over a silica gel column packed with 35–70 µm particles using hexane, DCM, ethyl acetate:hexane (1:3, v:v) and MeOH as eluents.

LCAs are contained in the second fraction (DCM) and were detected and quantified using gas chromatography with a flame-ionization detector (GC-FID). To remove compounds that can co-elute with LCAs such as alkenoates, LCA fractions were saponified by heating at 60 °C overnight in 1N KOH in MeOH:H₂O (95:5, v:v). After heating, the reaction mixture was cooled to room temperature, quenched with NaCl (5% by volume), extracted using hexane, and rerun on the GC-FID.

GC-FID analyses were performed on an Agilent 7890B Series GC system configured with an Agilent VF-200 ms capillary column (60 m length, 0.25 mm i.d., 0.10 µm film thickness) (Longo et al., 2013). Hydrogen was used as the carrier gas at a 36 cm/s column flow rate. The GC method used splitless injection (320 °C), and the oven temperature was programmed from 50 °C (hold for 1 min) to 255 °C at 20 °C/min, then to 300 °C at 3 °C/min, followed by 10 °C/min increase to 320 °C and hold for 10 min. Samples were

also run with the same temperature program on an Agilent 7890B Series GC coupled with a 5977A GC–EI mass spectrometer (GC–MS) to confirm the identity of the LCAs using the known ion chromatograms and by comparison of mass spectral data and GC retention times with published data (de Leeuw et al., 1980; Marlowe et al., 1984). LCAs were quantified using hexatriacontane (n -C₃₆ alkane) as an internal standard added to the sample before injection. The reproducibility achieved for duplicate LCA quantifications was estimated to be better than $\pm 10\%$ (coefficient of variation). The alkenone unsaturation index U_{37}^K and other indices (for example $U_{37}^{K'}$, $U_{37}^{K''}$, U_{38}^K) were calculated for all lakes following Brassell et al. (1986), Pearson et al. (2008), and Zheng et al. (2016). LCA distribution parameters such as C_{37}/C_{38} (Volkman et al., 1995), $\%C_{37:4}$ (Rosell-Melé, 1998), and RIK_{37} (Longo et al., 2016) were also computed.

2.3. Statistical analyses

Approximately 45% of the lakes sampled had LCAs below detection limit and where present, LCAs showed a strong right-skewed distribution. Consequently, the relationships between LCAs and environmental parameters were modelled using a two-stage hurdle process (Zuur et al., 2012) that models the presence-absence of LCAs and LCA concentration as two distinct processes. For the first part, or “hurdle”, we used a binomial generalised linear model based on the full dataset to model the presence and absence of LCAs above/below the detection limit. For observations with LCAs above the detection limit (i.e., those that crossed the hurdle), we modelled LCA concentration as a function of selected environmental variables using a log-normal model. The hurdle model considers presence/absence and concentration of LCAs separately, which is appropriate as environmental factors controlling LCA presence may be different from those controlling LCA concentration once observed (Taranu et al., 2017). Limnological datasets such as ours contain large numbers of potential predictors which are often highly intercorrelated. Variable selection is often used to find the “minimum adequate model” that contains a small subset of variables that explains, in a statistical sense, the data as well as the full set. However, variable selection with highly correlation predictors is problematic (Whittingham et al., 2006), and it is increasingly recognised that there is no “best” model, but often a series of models that explain the data equally well.

Predictive models were developed to account for the observation that many limnological variables are potentially correlated. For both parts of the hurdle model, we tested the significance of each variable (water chemistry, limnological, climatic, landscape) separately using a chi-squared or F test for the presence-absence and concentration models, respectively. We then fitted a large set of models using all combinations of the subset of significant variables (Calcagno and Mazancourt, 2010). Using this set of models, we then used Akaike's Information Criterion (AIC) to identify: (a) the best model, and (b) all other models within 2 AIC units of the best model, as these are effectively indistinguishable from the best model and are equally plausible. We then fitted a final model using the list of predictor variables that appear in any of the selected models.

Variance partitioning of either the variance explained for the log-normal model or pseudo- r^2 (binomial model; Nagelkerke, 1991) was used to quantify the importance or explanatory power of each predictor in the final model and to decompose the explained variance into independent and shared components (Borcard et al., 1992). To provide an additional visualisation of the relationship between LCAs and environmental factors, we used classification and regression trees (CARTs) to model LCA presence and absence, and concentration, respectively. CARTs are binary recursive tree-based models that identify a set of decision rules that best predicts either a binary outcome (classification trees) or quantitative variable (regression tree). At each stage of fitting a CART tree, the algorithm identifies a variable and threshold value that best predicts the response variable. Since our interest here is in explanation rather than prediction, we used AIC to identify the best tree rather than cross-validation (Hothorn and Zeileis, 2015). CARTs have the disadvantage that they can only display a single model, when there may be several best models, but have the advantage that they can reveal interactions between variables not easily visualised using other methods (Hastie et al., 2009).

The relationships between various alkenone-based indices ($\%C_{37:4}$, C_{37}/C_{38} ; U_{37}^K , $U_{37}^{K'}$, $U_{37}^{K''}$), alkenone isomer composition, and environmental factors were further explored using linear regressions and constrained ordinations. For the latter, we assessed the significance and importance of each predictor with a Monte Carlo permutation test and variance partitioning respectively (Borcard et al., 2011).

The distribution of LCA concentration and many of the explanatory variables showed a right-skewed distribution: these were either \log_{10} (all water chemistry, lake area, LCA concentration) or square-root- (sample depth) transformed prior to analysis to normalize their distributions. We also included average air temperature, obtained from the WorldClim database with a spatial resolution of approximately 1 km².

All statistical analyses were performed using R software for statistical computing and graphics (R Development Core Team, 2017) with the following packages: glmulti (Calcagno and Mazancourt, 2010), vegan (Oksanen et al., 2017), and partykit (Hothorn and Zeileis, 2015).

3. Results

3.1. Environmental data

Survey lakes exhibited large gradients for most environmental parameters (Table 1 and Supplementary Table S1) including depth (0.1–28.5 m), salinity (0.1–102.2 g/L) and mean water temperature (8.6–23.7 °C). In general, individual lakes had higher concentrations of sulfate (SO_4^{2-}) (31.5–120,223 mg/L) than bicarbonate (HCO_3^-) (156–5625 mg/L), carbonate (CO_3^{2-}) (<0.01–5528 mg/L) or chloride (Cl^-) (3.1–45,022 mg/L). In addition, lakes exhibited different degrees of thermal and chemical stratification, ranging from polymixis to permanent stratification (meromixis). Chemistry and hydrology of about two-thirds of these lakes has been discussed in Pham et al. (2009). Elevated sulfate concentration in the Saskatchewan region is mostly derived from oxidation of the underlying bedrock (Last and Ginn, 2005; Hayashi and van der Kamp, 2009).

3.2. LCA occurrence and abundances

Of the 106 lake surface sediments examined in this study, 58 (55%) contained detectable concentrations of LCAs (Fig. 1; Table 1). Total concentrations of LCAs range from 2.5 to 27,625 $\mu\text{g/g}$ TOC (0.02–2,312 $\mu\text{g/g}$ dry sediment), with very high concentrations (> 1000 $\mu\text{g/g}$ TOC or 100 $\mu\text{g/g}$ dry sediment) recorded in

nine of the LCA-containing lakes (15% of basins) (Table 1). The highest concentrations of LCAs generally occurred in the northern section of the sampling area ($> 51^{\circ}\text{N}$; Fig. 1).

The majority of LCA-containing surface sediments (53%) featured high $\%C_{37:4}$, with values $> 60\%$. Five lakes featured a profile mainly composed of the $C_{37:3}$ alkenone ($\%C_{37:4} < 40\%$) (Fig. 2). In addition, one lake displayed a predominance of C_{38} alkenones over C_{37} alkenones, with particularly elevated concentrations of the $C_{38:3}$ alkenone (Fig. 2). Among the lakes, four of them showed the presence of the $C_{37:3b}$ alkenone isomer, including one freshwater (< 0.5 g/L) and three oligohaline (0.5–5 g/L) lakes (Table 1 and Fig. 2). RIK_{37} (ratio between $C_{37:3a}$ and $C_{37:3b}$ isomers; Longo et al., 2016) values were in the range 0.47–0.57 (Table 1). C_{37}/C_{38} ratios varied between 0.4 and 7.7, with a mean value of 2.7 ± 1.7 (Table 1). No significant correlation was found between the U_{37}^K unsaturation index and depth-integrated water column temperature or surface water temperature at the time of sampling (August 2013), or with air (spring or summer) temperature. No significant correlation was found using other indices (such as the $U_{37}^{K'}$, $U_{37}^{K''}$, or the U_{38}^K), or through regression analysis of individual LCAs with air (spring or summer) and water temperature.

3.3. Factors influencing LCA presence and concentration

Fig. 3 shows the significant predictor variables explaining the presence/absence of LCAs, the explanatory power (variance) of each variable, and the explained variance decomposed into independent and shared components. Salinity, water pH, depth, water temperature and stratified (stratification of the lake) were the most significant predictor variables explaining the occurrence of LCAs in the Canadian prairie lakes (Supplementary Table S2). Salinity was the most important variable (24% of variance explained), and most of the variation explained by salinity (20%) was independent of other variables. Water pH explained a small part (3.5%), but its explanatory power was mostly independent of other parameters. Stratified, depth and water temperature explained moderate amounts of variability (5%, 11% and 14%, respectively), but their explanatory powers were largely confounded with each other.

To provide an additional visualisation of the relationship between LCAs and environmental factors, we used a classification tree (Fig. 4). A split in salinity at 2.4 g/L was observed, representing a threshold for the

presence of LCAs. At low salinity (< 2.4 g/L), stratification was important for the presence of LCA, but LCAs were rare in lakes with high pH (> 8.5). At higher salinity (> 2.4 g/L), LCAs were rare in shallow lakes (< 0.6 m). LCAs occurred mainly in deeper lakes (> 0.6 m), but they were slightly less common in very saline lakes (> 44.4 g/L).

Salinity, depth, water temperature and presence of stratification were the most significant predictor variables of modelled LCA concentrations (Fig. 5 and Supplementary Table S2). Salinity was the most important variable (31% of variance explained), and most of the variation explained by salinity was independent of other variables. Depth explained a much smaller proportion of variance (7%) and its explanatory power was entirely confounded with other variables. Stratification and water temperature explained moderate amounts (16% and 24%, respectively), although their explanatory powers were largely confounded with each other. The regression tree (Fig. 6) showed a salinity threshold of 4.9 g/L. A further distinction was noted at low salinities (< 4.9 g/L), with slightly higher LCA concentrations between 0.8 and 4.9 g/L than in less saline waters. At higher salinities (> 4.9 g/L), LCAs showed higher concentrations in stratified lakes than in unstratified basins.

Relative (%) importance of carbonate and sulfate and metrics of total salinity were identified as the main predictors of alkenone isomer compositions, but the explained variances were generally low (8.2%, 5% and 2.5%, respectively). Most variation explained by carbonate (4.6%) was independent of other variables, whereas the explanatory power of sulfate (4.8%) and salinity (1.4%) was largely confounded with other variables.

Statistical analyses between various LCA-based indices and environmental factors showed a significant but weak relationship between % carbonate (among anions) and %C_{37:4}, U₃₇^K, U₃₇^{K''}, or C₃₇/C₃₈ (Supplementary Table S2). No significant relationship was found between salinity and %C_{37:4}, or between salinity and U₃₇^K, U₃₇^{K''}, or C₃₇/C₃₈ (Supplementary Table S2). For the U₃₇^{K'} index, no significant correlation was observed with any environmental variables.

4. Discussion

4.1. LCA distribution and haptophyte taxonomy

The majority of LCA-containing lakes showed high proportion of $C_{37:4}$ alkenone (Fig. 2), which is a common feature of lacustrine systems (e.g., Sun et al., 2004; D'Andrea et al., 2011; Longo et al., 2016). A large variation in C_{37}/C_{38} ratio values (0.5–7.7; mean 3.3) was observed in these lakes, but values were in the same range as those recorded for *Isochrysis galbana* and *Ruttnera lamellosa* (Marlowe et al., 1984; Rontani et al., 2004) and lakes within the US Great Plain sites (Toney et al., 2010). In contrast, five Canadian lakes were characterized by elevated $C_{37:3}$ alkenone content (Fig. 2), a profile typical of offshore marine environments (e.g., Prahl et al., 1988; Sikes and Volkman, 1993) and rarely reported in lacustrine sediments (e.g., Li et al., 1996; Theissen et al., 2005; Toney et al., 2010; Hou et al., 2016). These lakes present lower C_{37}/C_{38} ratio variations and values (1.4–2.4; mean value = 1.8) similar to those recorded for *E. huxleyi* (Marlowe et al., 1984), from Chinese lakes (Sun et al., 2004; Chu et al., 2005), and in US sites with $C_{37:3}$ -dominant profiles (Toney et al., 2010). Finally, a single lake displayed a predominance of C_{38} over C_{37} LCAs, with especially elevated levels of tri-unsaturated components. A few lacustrine studies have reported such a profile in alkaline, evaporitic or saline settings (Thiel et al., 1997; Sun et al., 2004; Chu et al., 2005; Pearson et al., 2008). The C_{37}/C_{38} ratio was low (0.4) for this particular lake and very similar to that of the Spanish lakes (mean value = 0.1) studied by Pearson et al. (2008). Pearson et al. (2008) suggested that salinity may play a role in LCA composition, with a salinity threshold of 40 g/L for C_{37} LCAs and C_{38} -dominated LCA profiles being characteristic of hypersaline lakes. In contrast, the sole basin with C_{38} LCAs reported here was not hypersaline (10 g/L) and we report that C_{37} LCAs were also present in one lake with salinity > 40 g/L (Table 1), suggesting that salinity, per se, is not the sole factor influencing the dominance of C_{38} alkenones.

Differences in LCA profiles among basins may reflect variation in the community composition of LCA-producing haptophyte species in the studied Canadian lakes. Studies in the US Great Plains (Toney et al., 2010, 2012; Theroux et al., 2010) showed the presence of two haptophyte species from Group II among and within saline lakes, with OTU7 haptophyte or Hap B producing mainly $C_{37:3}$ alkenones, while OTU8 haptophyte or Hap A produces predominantly $C_{37:4}$ alkenones. Part of the reason for this difference in

distribution can be attributed to the offset in the timing of their bloom, where Hap A blooms in the early spring during low temperatures (2–12 °C) and Hap B blooms nearly a month later with a temperature range of (12–18 °C) (Theroux, 2012). In addition, Toney et al. (2012) showed that Hap A species dominates LCA production in sulfate-rich lakes (sulfate:carbonate ratio SCR > 1), while production is dominated by Hap B in carbonate-dominated lakes with SCR < 1. Here, the majority of the study lakes were sulfate-rich with SCR values much greater than one, except for Reflex Lake (Table 1 and Supplementary Table S1). All together, these observations would suggest that Hap A and Hap B are mainly responsible for the sedimentary LCA profile in the Canadian lakes, with few exceptions.

The Group I phylotype also thrived in some of the Canadian freshwater (< 0.5 g/L) and oligohaline (0.5–5 g/L) lakes, as suggested by Longo et al. (2016) based on the presence of the alkenone isomer C_{37:3b}. The RIK₃₇ index (ratio between C_{37:3a} and C_{37:3b} isomers) has also been proposed as a taxonomic indicator for multi-species effects, with RIK₃₇ values of 0.51–0.60 indicating lakes hosting only Group I haptophytes and RIK₃₇ values of 0.60–1.0 indicating lakes containing both Group I and Group II haptophytes (Longo et al., 2016). In this study, we found the C_{37:3b} isomer in four lakes (Fig. 2), including one freshwater and three oligohaline lakes, and RIK₃₇ values of 0.47–0.57 (Table 1), indicating the exclusive presence of Group I haptophytes in these lakes. In all other sites, RIK₃₇ values of 1 (C_{37:3b} isomer absent) suggest the exclusive presence of Group II haptophytes. Given these patterns, we will conduct genomic analyses in the future to confirm the presence of Hap A, Hap B, and Group I haptophytes in these Canadian lakes.

4.2. Relationship between LCAs and environmental parameters

Overall, our statistical analyses of the relationship between LCA distribution and major environmental gradients showed that LCA occurrence and abundance was influenced by a combination of salinity, water temperature, basin morphometry (depth), and water column stratification (Figs. 3–6). These results are consistent with previous studies that have shown that LCA abundance is generally higher in lakes with elevated sulfate content and salinity (e.g., Pearson et al., 2008; Toney et al., 2010; Liu et al., 2011; Zhao et al., 2014). Sulfate content does not appear as a main variable for LCA distribution in our statistical analyses,

likely because most of our study lakes are actually sulfate-rich. In contrast, salinity appears as a main factor on LCA presence and abundance. Salinity thresholds highlighted in our statistical analyses (2.4–44.4 g/L) support the observation that higher LCA concentrations occur in brackish/saline lakes rather than in fresh and hypersaline lakes (e.g., Liu et al, 2011; Song et al., 2016).

While salinity is the main variable regulating LCAs in this study, it was more difficult to separate the unique effects of water temperature (more LCAs with lower temperature), depth and stratification, likely because the thermal regime varied with the degree of lake stratification. In fact, highest LCA concentrations are generally recorded in lakes exhibiting permanent stratification and deep-water anoxia (Table 1). Stratification and deep-water anoxia have been suggested to play a role in LCA abundance in relation to the life cycle of lacustrine haptophytes (Toney et al., 2010). In the US Great Plains, the movement of LCAs within the water column is affected mainly by the degree of mixing and stratification experienced during the haptophyte bloom (Toney et al., 2010). Indeed, it appears that lacustrine haptophytes include a benthic vegetative stage within their life cycle (Toney et al., 2010), as recently shown for the marine haptophyte *Isochrysis galbana* (Ellegaard et al., 2016). In addition, some lacustrine phytoplankton cells survive during the winter season as resting cells in the bottom sediment (Salmi and Salonen, 2016). Thus, we conclude that bottom waters could represent a physical “refuge” for haptophyte cells during times of the year when conditions are unsuitable for population growth, such as winter.

In the Canadian Prairies, elevated sulfate content, derived from oxidation of pyrite in the underlying bedrock, is common in lakes that have significant groundwater contribution (Last and Ginn, 2005; Hayashi and van der Kamp, 2009). This environmental sulfur (as sulfate) is an essential element for all photosynthetic algae (e.g., Takahashi et al., 2011; Giordano and Prioretti, 2015); however, the influx of sulfate-rich groundwater could also favor chemical stratification (meromixis) and deep-water anoxia (Pham et al., 2008, 2009). Thus, at present, it is unclear how biotic demand and abiotic factors may interact to favour haptophytes and LCA production or preservation. In particular, further studies will be needed to further assess the influence of sulfate on LCAs and determine whether high LCA concentration in these Canadian lakes is the result of increased production or merely improved post-depositional preservation in the anoxic, aphotic bottom waters (Leavitt, 1993).

While the vast majority of our Canadian study lakes are saline, our survey also included 25 freshwater lakes (Table 1). Only 6 of these freshwater lakes contained detectable LCAs and our statistical analyses showed that LCAs occurred mainly in stratified alkaline freshwater lakes with pH up to 8.5 (Fig. 4). These observations are consistent with those in Alaskan freshwater lakes, where high LCA concentration was found in deep lakes with high pH, alkalinity and conductivity (Longo et al., 2016). Interestingly, these patterns suggest that the main environmental parameters influencing LCA occurrence and abundance may differ between freshwater and saline lakes, although it is still unclear how salinity, morphology, and stratification may interact with the basic biology of LCA-producing haptophytes.

4.3. Relationship between LCA-based indices and environmental parameters

The %C_{37:4} has been suggested as a salinity proxy as the relative proportion of the C_{37:4} alkenone has been reported to vary inversely with salinity in marine (e.g., Rosell-Melé, 1998; Schulz et al., 2000; Kaiser et al., 2017) and lacustrine environments (e.g., Liu et al., 2008, 2011; Song et al., 2016). As with some previous works (e.g., Sikes and Sicre, 2002; Toney et al., 2010), we found no significant relationship of %C_{37:4} with salinity (Supplementary Table S2), possibly because the production of C_{37:4} alkenone varies among haptophyte species and strains (Theroux et al., 2010). The U₃₇^K and U₃₇^{K''} indices, which include the C_{37:4} alkenone in their definition, are also not influenced by salinity variation (Supplementary Table S2), conversely to what has been observed in Chinese lakes (Liu et al., 2011; Song et al., 2016). On the other hand, a significant but weak correlation was observed between the relative abundance of C_{37:4} alkenone and relative importance of carbonate among anions (Supplementary Table S2). Similarly, a significant but weak correlation was found between U₃₇^K, U₃₇^{K''} and the % carbonate (Supplementary Table S2). The effect of carbonate on the proportion of C_{37:4} alkenone, if any, remains unknown.

4.4. Relationship between LCA unsaturation indices and temperature

Although temperature was a substantial (24%) predictor of variation in LCA concentrations in the statistical analyses of lake survey data, there was no significant relationship between mean water column temperature at the time of sampling (August 2013) and common indices of environmental temperature (U_{37}^K , $U_{37}^{K'}$, $U_{37}^{K''}$, or U_{38}^K). This lack of correlation may reflect the fact that temperature data was estimated on only a single date in August, while 1 cm deep sediment samples integrate historical haptophyte populations at the scale from seasons to years. Furthermore, late summer does not necessarily correspond to the time of maximum haptophyte production. For example, Toney et al. (2011) obtained a weak but positive and statistically significant correlation ($r^2 = 0.30$, $p = 0.067$), with average surface temperature recorded in spring in some of these lakes, suggesting a predominantly cool-season life cycle of haptophytes. The slope of the relationship was also similar to that derived from site-specific calibration of the U_{37}^K index in Lake George, North Dakota, suggesting that the change in LCA unsaturation as a function of temperature is similar between Great Plains lakes in the US and Canada. In fact, when we apply the temperature calibration of Lake George (Toney et al., 2010) at one site for which we have measured water temperature data for 20 years (Diefenbaker Lake), the LCA-inferred temperature is 11.9 °C, which is very close to the average water temperature in June (11.2 °C) and the average air temperature in the preceding month (May) recorded at a nearby weather station (10.9 °C; Elbow Cs weather station).

As the LCA profile at Lake Diefenbaker suggests the presence of Group I haptophyte, we also applied the three in situ temperature calibrations currently developed for lakes where the presence of Group I phylotype has been confirmed (D'Andrea et al., 2011, 2016; Longo et al., 2016). Here, the LCA-inferred temperature varied between 6.2 °C and 12.1 °C, values which were close to the average water temperature in May (6.8 °C) or June (11.2 °C). Together, these data support the idea that haptophyte blooms occur in spring after ice-out (Toney et al., 2010). Unfortunately, year-round estimates of water temperature were not available for the study lakes and we did not find any correlation with air temperature data for the spring season from weather stations close to the studied lakes.

Weak correlation between the LCA unsaturation indices and environmental temperature may also reflect the presence of multiple haptophyte species with different thermal optima (e.g., Hap A and Hap B), in a single lake. For example, Hap A produces mainly $C_{37:4}$ alkenones and blooms during spring, whereas Hap B

produces mainly $C_{37:3}$ alkenones and blooms during summer (Theroux, 2012). However, we did not find any significant correlations with spring air temperature for lakes with high % $C_{37:4}$ and with summer air temperature for lakes with low % $C_{37:4}$. This is likely because the temperature calibration throughout the bloom events, which combines Hap A and Hap B temperature calibrations, is actually very similar to the in situ lake calibration of Toney et al. (2010) (Theroux, 2012). Instead, we suggest that establishment of a valid calibration of environmental conditions and the U_{37}^K index may require more intensive field sampling during the haptophyte bloom in select candidate lakes to quantify the seasonality of LCA production and the appropriate environmental temperature for comparison (Toney et al., 2010).

4. Conclusions

Long-chain alkenones are common in the Canadian lakes of the northern Great Plains, and their presence and abundance is influenced by a combination of salinity, water temperature, depth, and degree of water-column stratification. Based on this study, sulfate may play a role in LCA distribution, even though it is unclear if its effect is related to biological requirements or to change in the aquatic environment (groundwater, stratification, etc.). While these environmental parameters clearly play a role in regulating the life cycle of the LCA-producing haptophytes, further studies will be essential to define how haptophyte ecology varies with seasonal and inter-annual changes in lake conditions. LCA profiles suggested that most lakes contained haptophyte species from the Group II phylotype, likely closely related to OTU8 haptophyte (Hap A) and OTU7 haptophyte (Hap B) found in the US Great Plains. However, while the presence of other species from the Group I phylotype was also confirmed, we note that they were not particularly common. Unexpectedly, we were not able to calibrate the LCA unsaturation ratios (U_{37}^K , $U_{37}^{K'}$, $U_{37}^{K''}$ or U_{38}^K) with environmental conditions at this time. Instead, further studies with a more intensive sampling at different depths and seasons will be needed to calibrate the U_{37}^K index in situ with the appropriate environmental temperature. Genomic analyses will also be important to precisely determine the haptophyte species present in these Canadian lakes.

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Figure captions

Fig. 1. Map of the southern Saskatchewan, Canada showing the location of the studied lakes. Individual lakes are plotted as “+” (no LCAs) or circles scaled to LCA concentration (µg/gTOC). The different LCA profiles found in this study are indicated by a colour code (white: high %C_{37:4}; grey: C_{37:3}-dominated; black: C₃₈-dominated).

Fig. 2. Signature sedimentary LCA profiles for the Canadian lakes. The profile with high (> 60%) %C_{37:4} (A) is observed in the great majority of the studied lakes, while the C_{37:3}-dominated (B) and C₃₈-dominated (C) profiles are only found in five lakes and a single lake, respectively. Four freshwater/oligohaline lakes also exhibit profile typical of Group I haptophyte (D), with the presence of tri-unsaturated isomers.

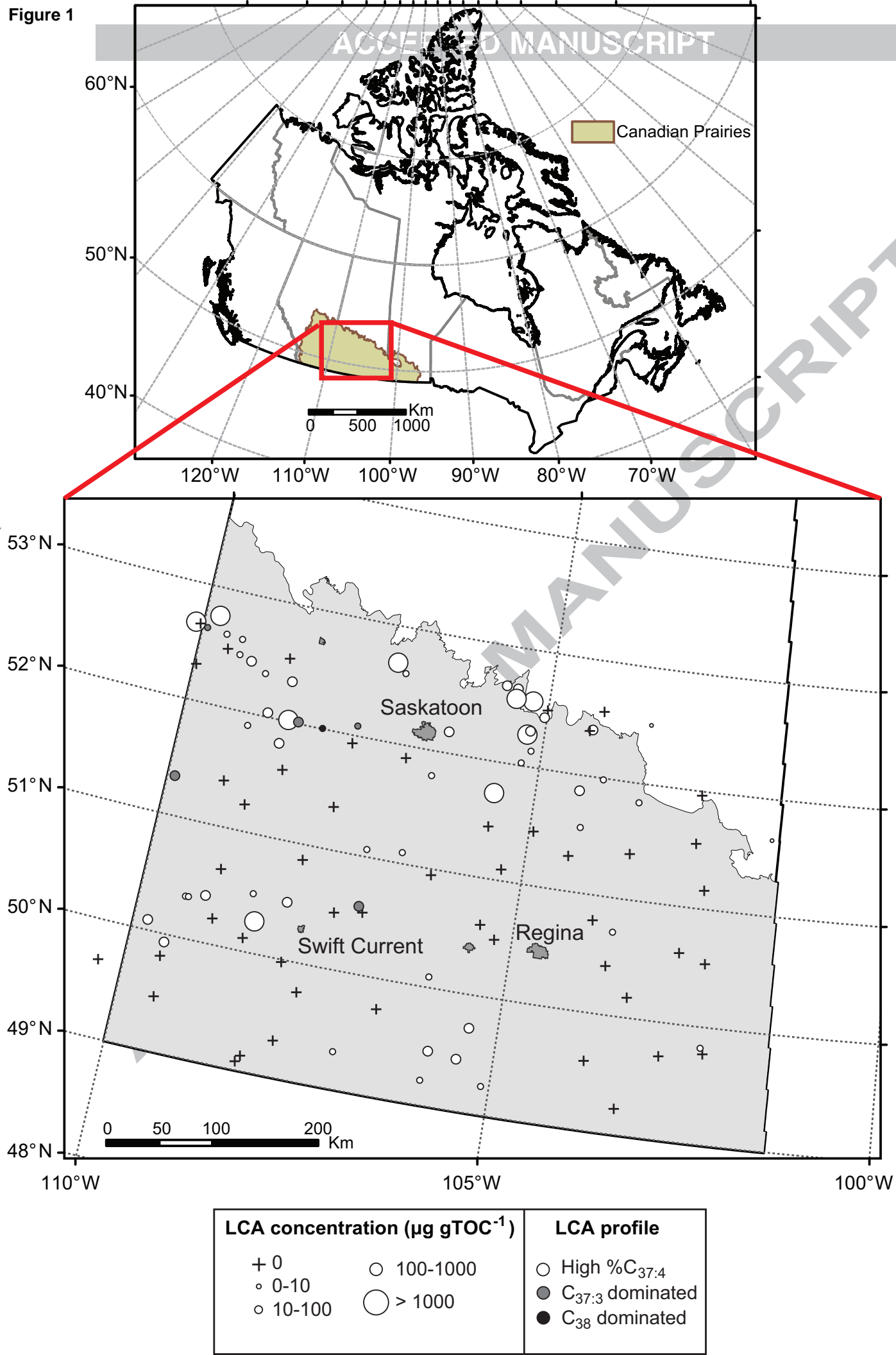
Fig. 3. Probability of LCA occurrence, showing (A)–(D) logistic (binomial model) regression plots, (E) mosaic plot of LCA occurrence vs thermal regime for raw data (left) and model fitted values (right), and (F) the shared and independent fraction of variance explained by each predictor in the final LCA model. Shaded areas show 95% CIs (confidence intervals) and ticks on the upper and lower axes indicate original observations. Statistics (r^2 , p value) for the regression plots (A)–(D) are presented in the Supplementary Table S2.

Fig. 4. Classification tree of LCA occurrence. Splits indicate thresholds for occurrence at particular environmental values. Bars at each node show proportions of observations where LCAs are present (dark grey) or absent (light grey).

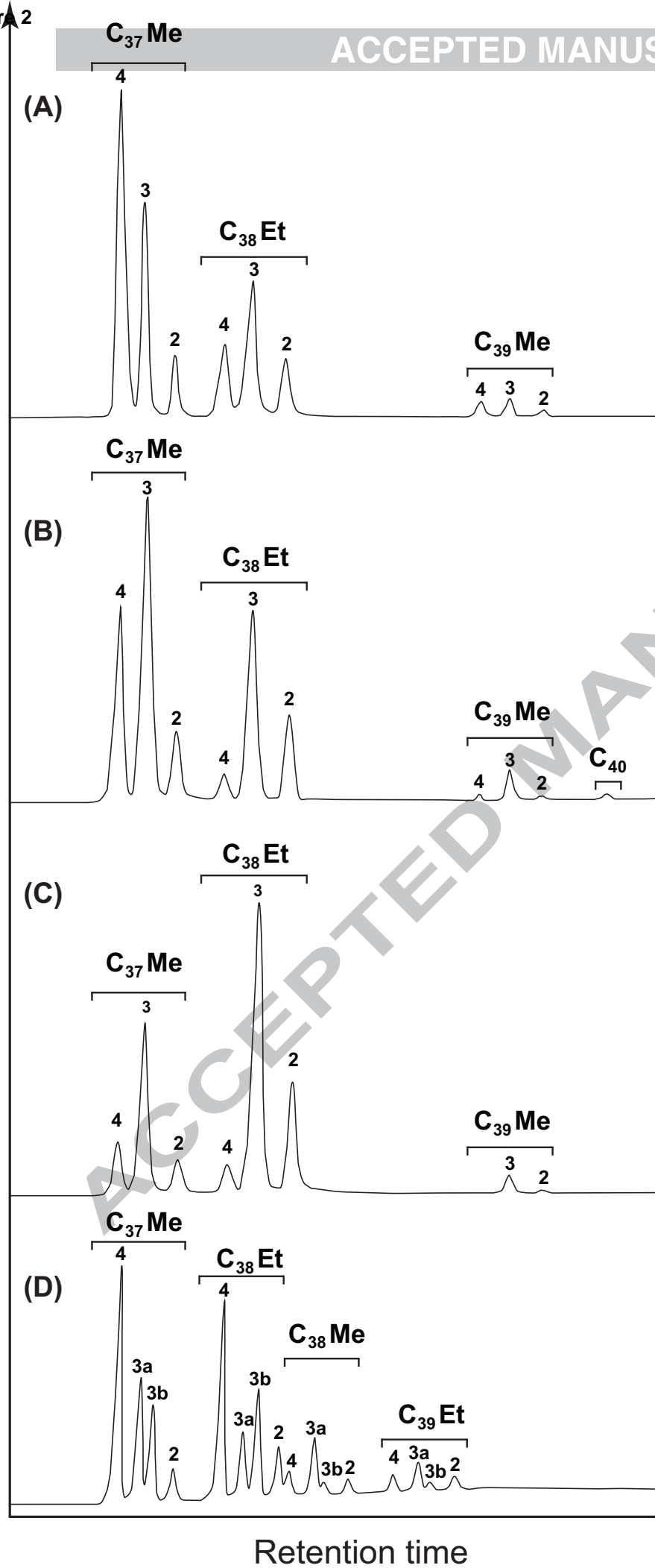
Fig. 5. LCA concentration as a function of selected significant environmental variables, showing (A)–(C) scatter plots and fitted regression lines (log-normal model), (D) boxplot of LCA concentration as a function of thermal regime, and (E) the shared and independent fraction of variance explained by each predictor in the final LCA model. Shaded areas show 95% CIs (confidence intervals) and ticks on the upper and lower axes indicate original observations. Error bars in (D) indicate model fitted values and 95% CIs. Statistics (r^2 , p value) for the scatter plots (A)–(C) are presented in the Supplementary Table S2.

Fig. 6. Regression tree of LCA occurrence. Splits indicate thresholds for occurrence at particular environmental values. Boxplots summarise LCA concentration for observations in that node. Note that boxplot scale is in log10 units (log10 of LCA concentration in $\mu\text{g/gTOC}$).

Figure 1



Response



Retention time

Figure 3

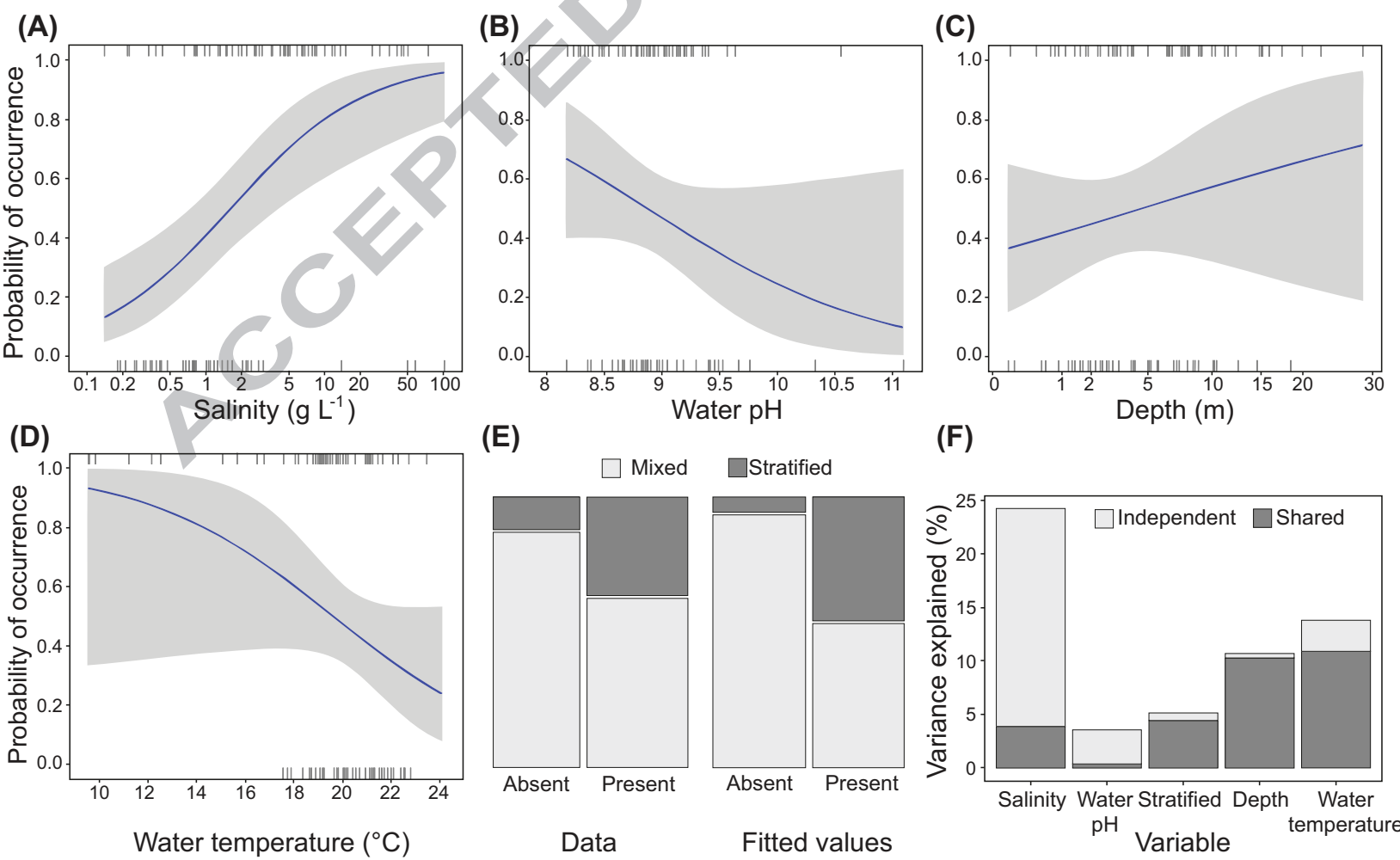


Figure 4

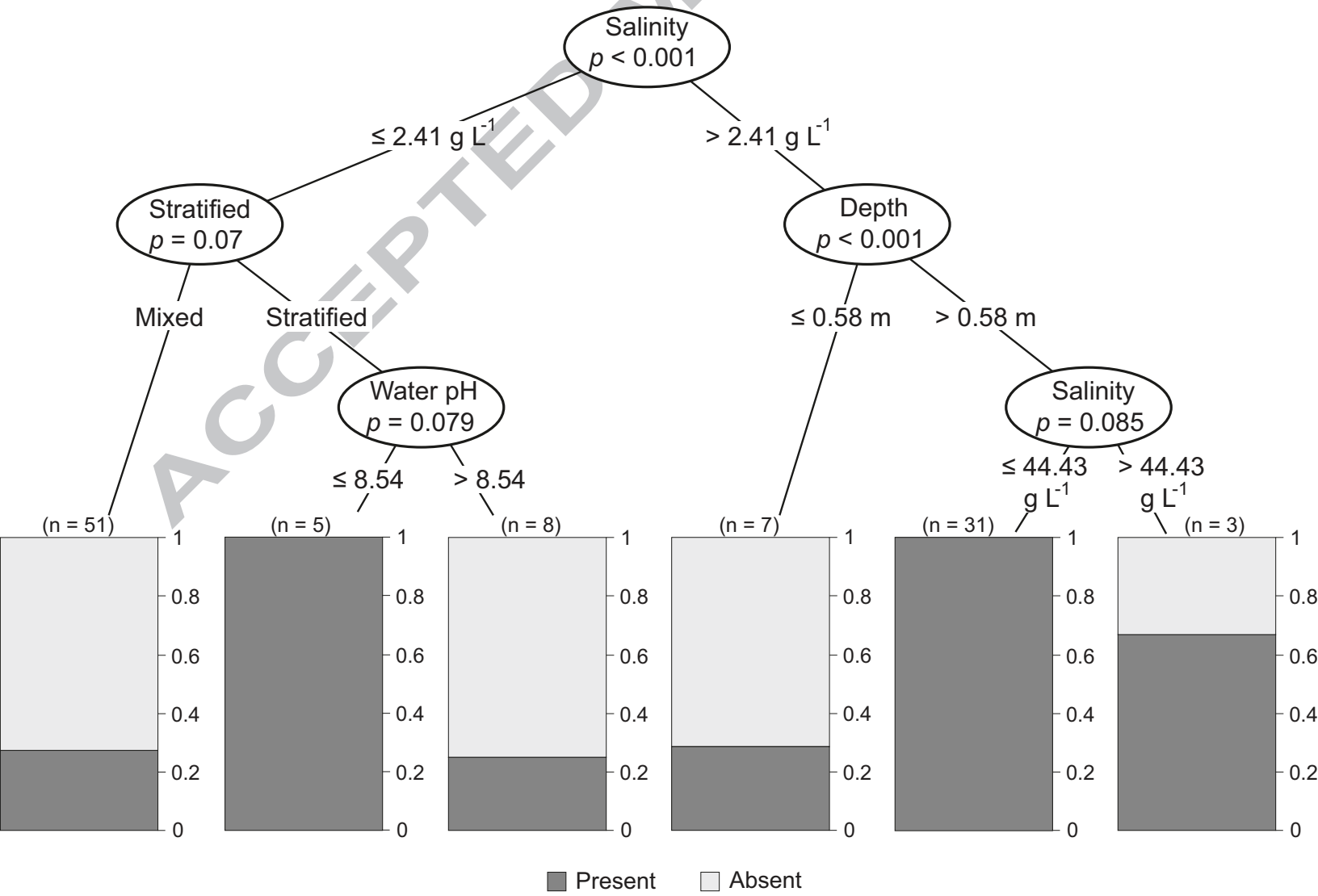


Figure 5

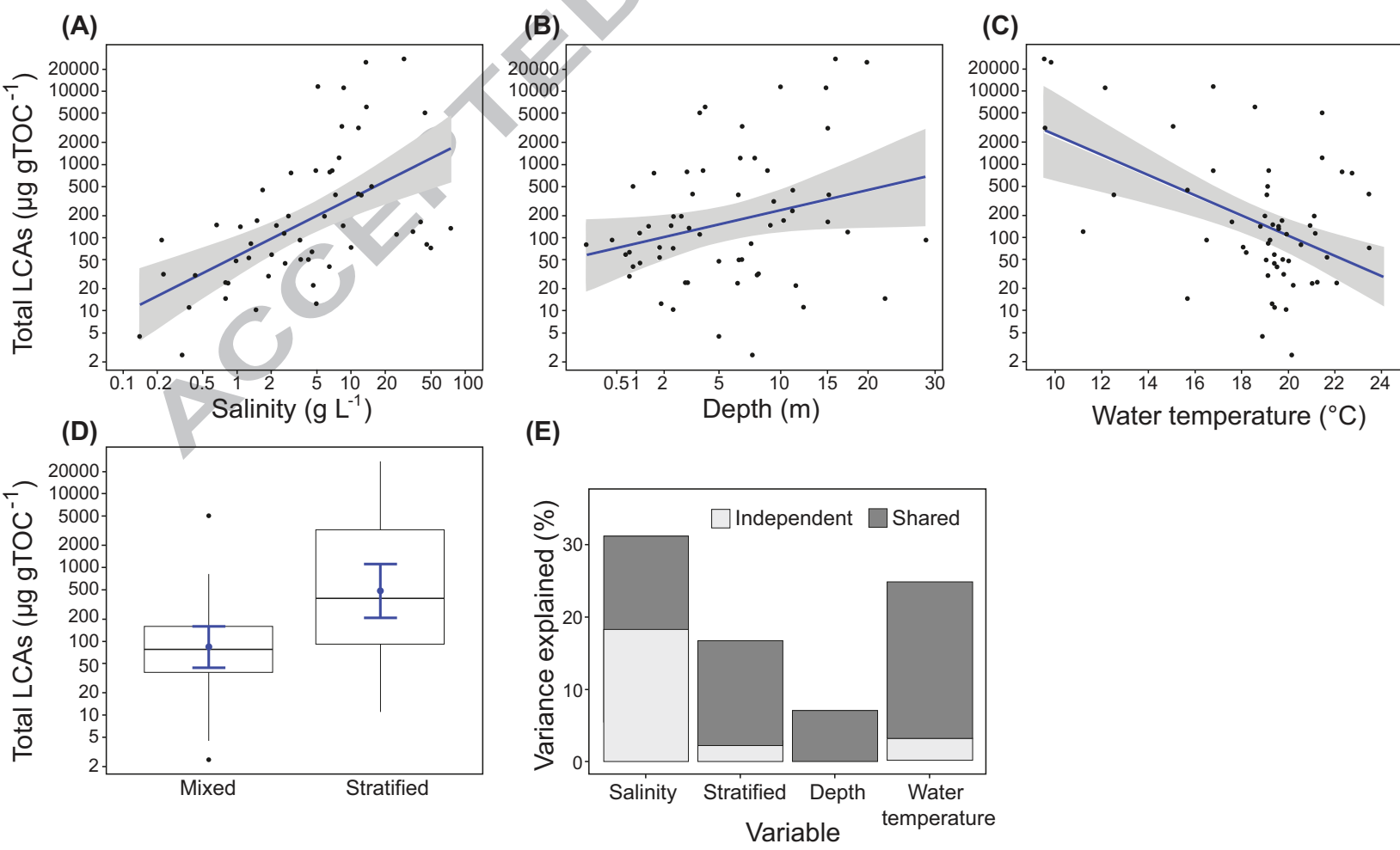
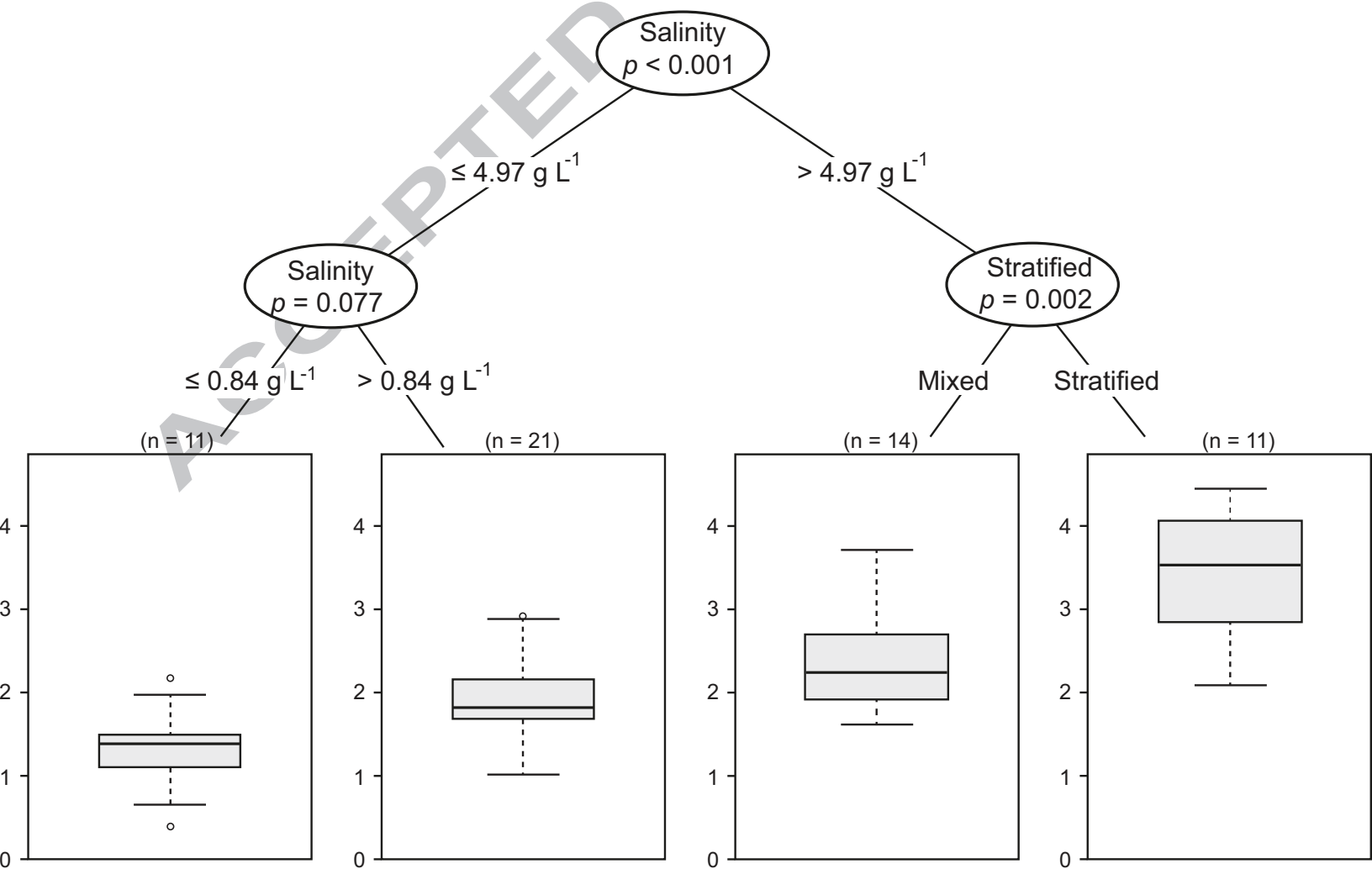


Figure 6



Lake Name	Lat. (°N)	Long. (°W)	Stratif. a	Depth (m)	Salinity (g/L)	Temp. (°C)	pH	SCR b	TOC (%)	LCAs (µg/gTOC)	LCAs (µg/gsed)	%C _{37:4} c	C ₃₇ /C ₃₈ d	RIK ₃₇ e	U ₃₇ ^K f	U ₃₇ ^{K'} g	U ₃₇ ^{K''} h
<i>LCAs present</i>																	
Alsask	51.34	109.88	N	3.2	74.8	19.6	9.35	17.7	3.21	135	4.33	29.6	1.59	1.00	-0.18	0.16	0.67
Antelope	50.28	108.39	N	7.7	7.87	21.4	9.40	91.1	7.89	799	63.0	70.5	5.17	1.00	-0.66	0.17	0.26
Arthur	52.56	105.44	T, M	6.7	8.29	15.1	8.88	N.D.	8.71	3296	287	69.9	2.83	1.00	-0.64	0.19	0.26
Bitter	50.10	109.79	N	0.9	15.1	19.1	8.69	510	2.15	609	13.1	46.9	1.28	1.00	-0.33	0.25	0.46
Blackstrap	51.79	106.44	N	7.9	0.43	19.1	8.84	47.2	8.22	26.9	2.21	52.0	2.59	1.00	-0.46	0.12	0.45
Bullkin	49.25	105.07	N	0.7	2.01	19.4	9.06	25.5	1.24	49.8	0.62	76.5	2.57	1.00	-0.70	0.26	0.18
Charron	52.40	104.31	T	8.8	4.9	16.9	10.5	125	14.6	927	135	58.2	2.82	1.00	-0.50	0.19	0.37
Clair	51.99	104.06	N	3.0	0.78	22.1	9.20	25.3	8.33	23.9	2.00	55.8	N.D.	1.00	-0.51	0.11	0.41
Coldspring	52.35	108.59	N	2.4	5.86	19.1	8.62	88.3	5.03	178	8.95	73.0	3.50	1.00	-0.62	0.40	0.18
Deadmoose	52.30	105.17	T, M	15.1	12.3	12.6	8.87	99.8	8.24	383	31.6	53.3	1.73	1.00	-0.41	0.27	0.39
Dewey	50.58	107.07	N	2.3	8.54	18.8	8.40	216	7.87	144	11.4	38.2	2.14	1.00	-0.26	0.19	0.57
Diefenbaker	51.10	106.62	N	28.5	0.22	19.3	8.27	19.8	3.11	87.6	2.72	52.1	1.38	0.47	-0.48	0.17	0.28
Eaglehill	52.36	108.97	N	1.1	2.64	18.7	9.03	1.92	4.61	37.5	1.73	79.5	N.D.	1.00	-0.76	0.17	0.18
Farewell	52.47	109.38	N	0.4	3.57	22.4	9.40	13.4	1.45	92.4	1.34	45.5	N.D.	1.00	-0.34	0.22	0.48
Fife	49.22	105.86	N	2.4	1.47	19.9	9.06	4.31	2.88	10.4	0.30	72.4	2.56	1.00	-0.63	0.33	0.20
Fishing	51.82	103.47	N	11.5	1.31	20.1	9.17	66.2	12.3	39.9	4.90	50.9	1.29	0.44	-0.45	0.23	0.27
Freefight	50.40	109.11	T, M	15	40.8	17.6	8.33	89.2	1.14	135	1.54	46.3	1.63	1.00	-0.29	0.32	0.44
Greenwater	52.52	103.50	N	7.5	0.33	20.1	8.23	153	17.2	2.48	0.43	43.3	N.D.	1.00	-0.23	0.36	0.46
Humboldt	52.14	105.12	N	6.3	0.84	20.8	9.13	31.8	9.89	23.7	2.34	55.5	7.05	1.00	-0.53	0.06	0.43
Junction	49.96	109.51	N	9.0	0.66	19.3	8.92	57.9	3.49	148	5.17	N.D.	N.D.	1.00	N.D.	N.D.	N.D.
Katepwa	50.71	103.65	T	22.4	0.8	15.6	8.29	237	6.38	14.7	0.94	47.7	6.35	1.00	-0.43	0.09	0.50
Kenosee	49.82	102.30	N	7.4	1.32	19.1	8.79	26.6	8.92	82.7	7.38	62.2	6.33	1.00	-0.59	0.08	0.36
L. Manitou	51.74	105.52	N	6.7	44.4	19.6	9.15	475	13.1	5063	662	55.6	2.24	1.00	-0.46	0.21	0.39
Lenore SB	52.45	105.00	T	9.1	4.43	21.1	9.38	39.9	16.1	143	22.9	39.4	0.88	0.54	-0.29	0.28	0.41
Little Manitou	52.62	109.62	N	3.8	3.62	21.5	9.19	4.56	13.1	50.1	6.55	48.7	4.00	0.49	-0.43	0.22	0.31
Luck	51.07	107.10	N	0.8	4.55	18.2	9.64	47.7	3.00	27.5	1.91	66.7	1.99	1.00	-0.57	0.28	0.26
Madge	51.65	101.63	N	12.3	0.38	19.4	8.36	235	8.61	11.0	0.95	51.3	1.66	1.00	-0.42	0.20	0.43
Manitou	52.76	109.75	T	15.9	29.1	8.75	9.56	13.2	8.37	27,575	2308	66.4	2.14	1.00	-0.59	0.22	0.28
Mason	50.36	109.35	N	1.9	4.97	19.3	9.25	21.2	5.02	12.4	0.62	72.3	3.04	1.00	-0.61	0.41	0.19

McLean	52.62	109.37	N	6.5	46.2	21.1	9.27	552	1.71	77.9	1.33	44.2	1.63	1.00	-0.39	0.10	0.53
Middle	52.56	105.19	T	10.0	5.14	16.8	8.93	88.7	6.40	11,522	737	72.9	1.29	1.00	-0.68	0.18	0.23
Montague	49.47	105.83	N	1.4	1.07	19.6	9.06	15.6	1.38	135	1.87	72.9	2.58	1.00	-0.67	0.22	0.22
Old Wives	50.10	105.99	N	1.7	1.51	18.9	9.29	6.78	0.10	17.9	0.02	N.D.	N.D.	1.00	N.D.	N.D.	0.33
Opuntia	51.81	108.57	N	1.6	2.98	21.4	9.02	35.6	2.41	603	14.5	75.8	7.73	1.00	-0.73	0.12	0.22
Porter	52.19	106.29	N	4.0	2.78	19.2	8.48	5.31	4.62	819	37.8	66.9	2.56	1.00	-0.59	0.25	0.27
Quill	51.86	104.37	N	6.4	7.29	19.1	8.78	189	8.05	383	30.8	65.1	4.13	1.00	-0.51	0.41	0.24
Rabbit	52.61	107.02	S	6.5	4.23	19.3	9.21	80.5	11.8	72.6	8.60	70.0	4.49	1.00	-0.64	0.19	0.26
Redberry	52.62	107.02	T	14.8	8.61	12.1	9.09	22.8	4.40	11,143	490	69.4	2.15	1.00	-0.64	0.16	0.27
Reesor	49.66	110.11	N	5.0	0.14	18.9	8.48	N.D.	9.15	10.3	0.41	72.5	1.64	1.00	N.D.	N.D.	0.28
Richmond	52.01	108.02	N	1.8	10.0	18.8	8.62	3634	12.1	73.3	8.88	34.1	0.38	1.00	-0.24	0.16	0.62
Shannon	52.65	105.43	T	11.2	1.67	15.8	9.06	53.4	9.53	445	42.4	61.6	2.80	1.00	-0.55	0.18	0.34
Shoe	49.72	105.36	N	3.8	25.1	19.9	8.90	266	6.94	110	7.64	41.4	1.71	1.00	-0.28	0.23	0.52
Sink	52.44	109.19	N	2.8	2.83	20.9	8.66	12.2	14.7	175	25.8	79.2	2.78	1.00	-0.76	0.18	0.18
Snakehole	50.50	108.49	N	2.4	50.2	23.2	8.96	N.D.	4.16	191	7.96	45.6	1.97	1.00	-0.38	0.13	0.51
Springwater	52.01	108.39	N	3.1	6.56	22.4	8.62	47.5	4.40	797	35.1	37.5	2.17	1.00	-0.21	0.26	0.55
Success	50.49	108.02	T, M	17.4	34.9	11.3	8.62	43.4	4.66	94.2	4.39	45.1	1.50	1.00	-0.30	0.28	0.47
Sufferin	52.64	109.90	N	8	0.22	19.7	8.18	N.D.	12.7	31.8	4.03	37.3	1.64	1.00	-0.28	0.14	0.59
Summercove	49.32	107.04	N	5	0.98	20.0	8.97	13.1	5.24	31.5	1.65	67.8	6.97	1.00	-0.65	0.09	0.30
Sunny	52.02	108.52	T, M	15	11.6	8.63	8.50	43.3	7.94	3143	249	63.5	2.27	1.00	-0.56	0.21	0.31
Tramping	52.03	108.82	N	3.4	11.6	23.7	9.14	121	12.6	99.3	12.5	67.3	0.49	1.00	-0.63	0.13	0.30
Unknown 1	51.55	104.27	N	1.8	1.27	21.7	8.73	N.D.	5.24	53.3	2.79	69.8	0.93	1.00	-0.60	0.32	0.23
Van Scoy	52.08	107.55	N	0.9	6.44	20.6	8.82	N.D.	7.03	17.6	1.24	28.9	1.40	1.00	-0.05	0.33	0.62
Wakaw	52.66	105.60	T	10.3	1.5	19.7	8.54	5.06	12.2	132	16.1	68.6	N.D.	1.00	-0.67	0.05	0.30
Waldsea	52.26	105.18	T, M	19.9	12.8	9.85	8.45	165	8.51	24,494	2084	71.9	2.81	1.00	-0.67	0.16	0.25
White Heron	51.89	109.05	N	0.8	1.88	21.9	8.94	12.7	6.46	29.5	1.91	67.7	N.D.	1.00	-0.62	0.17	0.28
Willow Bunch	49.45	105.46	N	1.1	2.58	21.1	9.16	19.1	1.30	108	1.40	76.8	3.37	1.00	-0.71	0.27	0.18
Wolverine	52.03	105.22	N	3.1	0.8	21.2	8.93	2.05	12.5	21.2	2.65	76.9	4.31	1.00	-0.67	0.44	0.14
X5	52.67	110.08	T	4.1	13.6	17.9	9.04	20.8	12.2	6064	741	65.1	2.91	1.00	-0.59	0.18	0.31

LCA's absent

Adrian	49.69	102.87	N	1.2	0.79	19.8	9.13	1.98	5.37	x ⁱ	x	x	x	x	x	x	x
Alioth	52.51	109.55	N	0.6	13.9	21.2	10.3	179	9.14	x	x	x	x	x	x	x	x

Annie Laurie	51.97	102.67	N	1.3	0.25	22.1	8.78	23.3	20.0	x	x	x	x	x	x	x	x
Barber	51.37	107.66	N	1.6	1.26	20.4	8.57	34.1	3.87	x	x	x	x	x	x	x	x
Buffalo Pound	50.61	105.43	N	4.1	0.31	22.2	8.85	1.27	4.94	x	x	x	x	x	x	x	x
Chapleau	50.18	103.34	N	1.4	0.41	22.1	9.00	15.3	9.75	x	x	x	x	x	x	x	x
Clearwater	50.87	107.93	N	7.0	0.68	19.1	9.18	15.8	9.27	x	x	x	x	x	x	x	x
Crooked	50.61	102.72	N	10.2	0.82	21.0	8.72	49.8	5.70	x	x	x	x	x	x	x	x
Cutbank	50.53	107.00	S	2.4	2.41	20.4	9.13	23.1	5.87	x	x	x	x	x	x	x	x
Cypress	49.47	109.47	N	5.6	0.3	19.1	8.82	115	2.85	x	x	x	x	x	x	x	x
Deep	50.42	103.68	N	2.2	1.53	21.8	8.88	62.9	6.56	x	x	x	x	x	x	x	x
Edouard	52.38	104.35	N	5.6	0.34	19.9	11.1	98.6	13.4	x	x	x	x	x	x	x	x
Elkwater	49.67	110.29	N	7.9	0.18	18.7	8.65	67.6	4.25	x	x	x	x	x	x	x	x
Eyeblow	50.96	106.19	N	0.9	0.21	17.9	8.36	N.D.	1.34	x	x	x	x	x	x	x	x
Good Spirt	51.55	102.67	S	5.2	0.47	19.8	8.62	1.22	11.2	x	x	x	x	x	x	x	x
Gouverneur	49.76	107.65	N	7.2	1.38	19.8	8.73	35.9	2.37	x	x	x	x	x	x	x	x
Grassy	51.24	104.63	N	2.0	0.64	21.2	8.98	9.40	12.6	x	x	x	x	x	x	x	x
Gull	50.11	108.50	N	5.0	2.04	20.1	8.96	38.3	N.D.	x	x	x	x	x	x	x	x
Handsome	50.49	107.38	N	1.9	2.19	18.4	9.04	28.1	6.89	x	x	x	x	x	x	x	x
Harris	49.82	109.50	N	5.1	0.35	18.9	8.83	133	0.86	x	x	x	x	x	x	x	x
Horse	51.39	103.56	N	3.3	0.72	20.7	8.78	8.20	22.1	x	x	x	x	x	x	x	x
Island	49.13	108.23	N	2.0	0.18	21.7	8.87	7.31	6.62	x	x	x	x	x	x	x	x
Kindersley	51.42	109.10	N	1.6	0.42	17.6	9.66	37.7	2.02	x	x	x	x	x	x	x	x
Kipabiskau	52.57	104.18	N	6.8	0.78	21.7	8.90	N.D.	16.6	x	x	x	x	x	x	x	x
Lac Pelletier	49.99	107.94	N	8.8	0.41	20.9	9.46	1.04	12.1	x	x	x	x	x	x	x	x
Last Mountain	51.10	105.26	N	18.5	1.05	19.2	8.84	1.80	9.25	x	x	x	x	x	x	x	x
Lenore	52.51	104.97	T	10.1	2.26	17.7	8.97	168	7.77	x	x	x	x	x	x	x	x
Lewis Creek South	51.44	105.53	N	1.9	1.36	21.1	8.74	N.D.	8.21	x	x	x	x	x	x	x	x
Lonetree	49.08	108.26	N	4.0	0.19	20.2	9.05	7.09	7.32	x	x	x	x	x	x	x	x
Macklin	52.32	109.95	N	2.8	0.8	20.2	8.38	N.D.	12.3	x	x	x	x	x	x	x	x
McDonald	49.19	103.24	N	14.5	0.25	20.0	8.48	N.D.	2.60	x	x	x	x	x	x	x	x
Newton	49.32	107.83	N	4.2	0.38	20.6	8.95	32.0	1.54	x	x	x	x	x	x	x	x
Nickle	49.59	103.78	N	5.7	0.25	19.2	8.17	37.6	2.27	x	x	x	x	x	x	x	x
Pasqua	50.79	103.94	T	12.5	0.77	21.2	8.78	23.7	5.59	x	x	x	x	x	x	x	x
Peter	51.46	104.91	N	0.5	2.78	22.4	9.49	165	12.6	x	x	x	x	x	x	x	x

Pike	51.90	106.82	N	2.5	0.26	19.6	8.66	2.47	12.8	x	x	x	x	x	x	x	x
Reflex	52.66	110.01	N	2.8	102	22.2	9.40	0.95	4.22	x	x	x	x	x	x	x	x
Round	50.54	102.37	N	10.4	0.83	21.4	8.82	26.6	4.31	x	x	x	x	x	x	x	x
Shrimp	51.59	108.45	N	3.0	0.41	22.6	9.05	23.9	11.8	x	x	x	x	x	x	x	x
Snipe	51.23	108.86	N	1.2	1.67	18.6	8.85	35.4	3.40	x	x	x	x	x	x	x	x
Stony Beach	50.50	105.19	S	2.7	0.35	22.1	9.53	1.08	6.00	x	x	x	x	x	x	x	x
Thackeray	52.54	108.69	N	1.6	1.17	19.6	9.42	24.7	8.17	x	x	x	x	x	x	x	x
Thomson	49.77	106.60	N	8.3	1.01	20.3	8.87	40.0	N.D.	x	x	x	x	x	x	x	x
Unknown 3	50.65	108.97	N	0.1	57.8	21.5	8.57	N.D.	0.23	x	x	x	x	x	x	x	x
Vincent	50.22	108.94	N	0.1	49.9	22.0	9.42	N.D.	2.61	x	x	x	x	x	x	x	x
WhiteBear	49.77	102.26	N	8.5	2.03	19.2	8.67	16.3	7.75	x	x	x	x	x	x	x	x
X3	51.94	107.58	N	0.6	3.08	19.8	9.76	457	7.34	x	x	x	x	x	x	x	x
York	51.16	102.48	N	3.0	1.09	22.8	8.97	21.6	14.2	x	x	x	x	x	x	x	x

^a Stratification: none (N), stratified (S), thermal (T), meromictic (M)

^b SCR: Sulfate to Carbonate Ratio (Toney et al., 2010)

^c %C_{37:4} = 100×([C_{37:4}]/[C₃₇ LCAs]) (Rosell-Melé, 1998)

^d C₃₇/C₃₈ = [C₃₇ LCAs]/[C₃₈ LCAs] (Volkman et al., 1995)

^e RIK₃₇ = [C_{37:3a}]/[C_{37:3a} + C_{37:3b}] (Longo et al., 2016)

^f U₃₇^K = [C_{37:2} - C_{37:4}]/[C_{37:2} + C_{37:3} + C_{37:4}] (Brassell et al., 1986)

^g U₃₇^{K'} = [C_{37:2}]/[C_{37:2} + C_{37:3}] (Brassell et al., 1986)

^h U₃₇^{K''} = [C_{37:3}]/[C_{37:3} + C_{37:4}] (Zheng et al., 2016)

ⁱ x = not detected

Highlights

- Alkenones (LCAs) were investigated in 106 lakes in the Canadian Prairies
- LCAs present in 55% of surveyed lakes, with very high concentrations in 7% of lakes
- Salinity and stratification play key roles for LCA presence and abundance
- Improved seasonal sampling needed to better identify the timing of LCA production